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## Useful Synthesis of the Main Dehydrohexapeptide Segment of a Macrocyclic Antibiotic, Berninamycin B

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The useful synthesis of tetradehydrohexapeptide segment **2**, which is the main skeleton of a macrocyclic antibiotic, berninamycin B, was first achieved. The skeleton **2** is constructed, in turn, of  $2-[(Z)-1-amino-1-propenyl]-5-methyl-oxazole-4-carboxylic acid, <math display="inline">\alpha$ -dehydroalanine ( $\Delta Ala$ ), L-Val, 2-[1-amino-1-ethenyl]-5-methyloxazole-4-carboxylic acid residues, besides L-Thr and  $\Delta Ala$  at the N- and C-termini, respectively.

Antibiotic berninamycin B (1),<sup>1,2</sup> isolated from the culture of *Streptomyces bernensis*, is a unique macrocyclic polyoxazole dehydropeptide. The natural product (1) features two interesting substructures, the main dehydrohexapeptide segment 2 called the Fragment B-C and the central heterocyclic skeleton 3 called the Fragment A, the former of which contains three vinyl and one 1-propenyl functional groups, as shown in Figure 1.

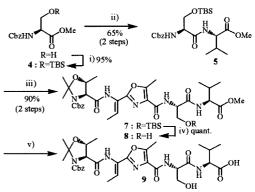
Figure 1. Berninamycin B (1).

Recently, we have reported briefly on the novel syntheses of partial Fragment B<sup>3</sup> and pyridine skeleton (3),<sup>4</sup> the latter of which is the common structure of similar antibiotics A10255 G and J.<sup>5</sup> The attractive structure as well as the bioactivity of 1 prompted us to study the total synthesis and the structure–bioactivity relationship.

Herein, we wish to report convenient synthesis of **2** from the two building blocks, the N,O-protected dehydrotetrapeptide **9** [carboxy (C-) component] and the N,O-protected dipeptide **18** [amine (N-) component]. Furthermore, final fragment condensation between the C- and N-components and then stepwise  $\beta$ -elimination of the three OH groups of the Ser residue were first achieved to give the desired N,O-protected tetradehydrohexapeptide **2** [(**P**)-**2**].

At first, to synthesize **9**, *N*-benzyloxycarbonyl(Cbz)-Ser-OMe was protected with *t*-butyldimethylsilyl chloride (TBSCl) to give the corresponding Ser(TBS) derivative **4**. Ester hydrolysis with 1 M LiOH in dioxane, followed by coupling with L-Val-OMe gave Cbz-Ser(TBS)-L-Val-OMe (**5**) in two steps. Subsequently, after deprotecting the Cbz group of **5** with 10% Pd-C, fragment condensation with 2-{*N*-Cbz-*N*,*O*-isopropylidene(Ip)-L-Thr-[(*Z*)-1-amino-1-propenyl]}-5-methyloxazole-4-

caboxylic acid (6) <sup>3</sup> as the *C*-component by using BOP<sup>6</sup> as a condensing agent and (*i*-Pr)<sub>2</sub>NEt gave the corresponding dehydrote-trapeptide methyl ester **7** containing Ser(TBS) residue. Deprotection of TBS group with 1 M tetrabutylammonium fluoride (TBAF) gave the expected 2-{*N*-Cbz-*N*,*O*-Ip-L-Thr-[(*Z*)-1-amino-1-propenyl]}-5-methyloxazole-4-carbonyl-L-Ser-L-Val-OMe (8). <sup>7</sup> Ester hydrolysis of **8** with 1 M LiOH gave the corresponding Fragment B derivative **9**, which was immediately subjected to the later condensation, without purification, as shown in Scheme 1.



Scheme 1. Reagents and conditions: i) TBSCl, imidazole, DMF, 0 °C, 1 h, rt, 1 h, ii) (a) 1 M LiOH, H<sub>2</sub>O-dioxane, 0 °C, 1 h, rt, 3 h, (b) DCC, HOBt, DMF, H-Val-OMe, 0 °C, 1 h, rt, 6 h, iii) (a) 10% Pd-C, H<sub>2</sub>, MeOH, rt, 3 h, (b) 6, BOP, (*i*-Pr)<sub>2</sub>NEt, 0 °C, 1 h, rt, 6 h, iv) 1 M TBAF, THF, rt, 1 h, v) 1 M LiOH, H<sub>2</sub>O-dioxane, 0 °C, 1 h, rt, 3 h

On the other hand, to synthesize the precursor of Fragment C derivative 18, coupling of N-Cbz-N,O-Ip-L-Ser-OH (10), derived from L-Ser for 4 steps, with L-Thr(TBS)-OMe by using DCC and HOBt, followed by deprotection of the TBS group with 1 M TBAF gave the corresponding Ser-Thr-OMe (11). Intramolecular cyclization of 11 by the oxidations with Jones reagent and then with PPh3 and I2 proceeded to give the corresponding 5-methyloxazole-4-carboxylate 13 via unstable αacetyldipeptide intermediate 12. Subsequently, firstly, ester hydrolysis of 13 with 1 M LiOH, followed by elongation of the obtained carboxylic acid 14 with H-Ser(TPS)-OMe (TPS = tbutyldiphenylsilyl) by using diphenylphosphoryl azide (DPPA) in the presence of Et<sub>3</sub>N gave the corresponding 5-methyloxazole-4-carbonyl-Ser(TPS)-OMe 15. Secondarily, selective cleavage of the Ip group of 15 with trifluoroacetic acid (TFA) gave 2-(1-amino-2-hydroxy)ethyl-5-methyloxazole derivative 16, the OH group of which was protected with triethylsilyl chloride (TESC1) to give 2-[1-N-(Cbz)amino-2-O-(TES)hydroxy]ethyl-5-methyloxazole derivative 17.8 Finally, hydrogenolysis of Cbz group of 17 gave the expected N-deprotected dipeptide 18 as the precursor of the N-component of 2, as shown in Scheme 2.

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Scheme 2. Reagents and conditions: i) (a) CbzCl, NaHCO $_3$ , Et $_2$ O-H $_2$ O, 0 °C, 2 h, rt, 4 h, (b) CH $_3$ I, KHCO $_3$ , DMF, 0 °C, 1 h, rt, 6 h, (c) DMP, TosOH, acetone, rt, 12 h, (d) 1 M LiOH, H $_2$ O-dioxane, 0 °C, 1 h, rt, 3 h, ii) (a) H-Thr(TBS)-OMe, DCC, HOBt, DMF, 0 °C, 1 h, rt, 6 h, (b) TBAF, THF, rt, 6 h, iii) Jones reagent, acetone, 0 °C, 1 h, rt, 2 h, iv)  $_1$ , PPh $_3$ , Et $_3$ N, CHCl $_3$ , 0 °C, 30 min, v) 1 M LiOH, MeOH, vi) H-Ser(TPS)-OMe, DPPA, Et $_3$ N, DMF, 0 °C, 1 h, rt, 6 h, vii) TFA, CHCl $_3$ , 0 °C, 30 min, rt, 1 h, viii) TESCl, imidazole, CH $_2$ Cl $_2$ , 0 °C, 1 h, rt, 1 h, ix) 10% Pd-C, H $_2$ , EtOH, rt, 1 h

Furthermore, coupling of 9 with 18 by the BOP method gave the expected  $\emph{O}\text{-TES}$  protected dehydrohexapeptide 19, which was then subjected to the simultaneous  $\beta$ -elimination of the three hydroxy groups. At present, however, the satisfactory results have not been obtained yet. Therefore, the stepwise elimination was tried successfully to give the expected 2 as follows.

Scheme 3. Reagents and conditions: i) BOP,  $(i\text{-Pr})_2\text{NEt}$ ,  $\text{CH}_2\text{Cl}_2$ , 0 °C, 1 h, rt, 12 h, ii) (a) MsCl,  $\text{Et}_3\text{N}$ ,  $\text{CHCl}_3$ , 0 °C, 30 min, (b) DBU, CHCl<sub>3</sub>, 0 °C, 30 min, (iii) 70% AcOH, THF, rt, 30 min, iv) (a) MsCl,  $\text{Et}_3\text{N}$ , CHCl<sub>3</sub>, 0 °C, 30 min, (b) DBU, CHCl<sub>3</sub>, 0 °C, 30 min, v) TBAF, THF, 0 °C, 1 h, rt, 3 h, vi) (a) MsCl,  $\text{Et}_3\text{N}$ , CHCl<sub>3</sub>, 0 °C, 30 min, (b) DBU, CHCl<sub>3</sub>, 0 °C, 30 min, (b) DBU, CHCl<sub>3</sub>, 0 °C, 30 min

The central Ser residue was converted to  $\alpha$ -dehydroalanine ( $\Delta$ Ala) by successive mesylation with MsCl and  $\beta$ -elimination with DBU to give didehydrohexapeptide **20** by the method reported earlier. After selective deprotection of the TES group with 70% AcOH, the OH group of the formed **21** was further mesylated and then  $\beta$ -eliminated to give the corresponding tridehydrohexapeptide **22**. Finally, similarly to the case of **21**, deprotection of TPS group of **22** with TBAF gave tridehydrohexapeptide **23**, followed by  $\beta$ -elimination gave the expected main skeletal tetradehydrohexapeptide **2**, as shown in Scheme 3.

The structures of all the new products thus obtained were confirmed by the  $^1H$  NMR spectral data and the satisfactory results of the elemental analyses. In particular, from the  $^1H$  NMR spectrum of 2, the chemical shifts of six protons of the three vinyl group at  $\delta = 5.45$ , 5.77, 5.95, 6.45, 6.56 and 6.69 ppm and that of olefinic proton of the propenyl group at  $\delta = 6.56$  ppm were found to be very similar to those of the natural berninamycin B (1).

In conclusion, a convenient synthesis of the main tetradehydrohexapeptide segment 2 of 1 was first accomplished. By using this result, further investigation on the total synthesis of 1 is currently under way in our laboratory.

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## References and Notes

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- 5 a) L. D. Boeck, D. M. Berry, and R. W. Wetzel, J. Antibiot., 45, 1222 (1992). b) M. Debono, R. Molly, J. L. Occolowitz, J. W. Paschal, A. H. Hunt, K. M. Michel, and J. M. Martin, J. Org. Chem., 45, 1809 (1992).
- 6 BOP: Benzotriazol-1-yloxy-tris(dimethylamino)phosphonium hexafluorophosphate.
- **8**: Colorless syrup.  $[\alpha]_{0}^{27}$  +4.45° (*c* 1.03, MeOH). <sup>1</sup>H NMR(CDCl<sub>3</sub>) δ = 0.88, 0.90(each d, 6H, Val's CH<sub>3</sub> × 2, J = 8.5, 9.0 Hz), 1.51(d, 3H, Thr's CH<sub>3</sub>, J = 6.0 Hz), 1.56–1.85(m, 9H, Ip and propenyl's CH<sub>3</sub>), 2.18(m, 1H, Val's β-H), 2.60(s, 3H, oxazole's CH<sub>3</sub>), 3.47(s, 3H, OCH<sub>3</sub>), 4.05(d, 1H, Thr's α-H, J = 6.0 Hz), 4.14(m, 1H, Ser's α-H), 4.35(m, 1H, Thr's β-H), 4.50(d, 1H, Val's α-H, J = 8.5 Hz), 4.60(m, 1H, Ser's α-H), 5.13(m, 2H, Cbz's CH<sub>2</sub>), 6.50(br s, 1H, propenyl's H), 7.19–7.27(m, 6H, Cbz's Ph and NH), 7.48(br s, 1H, NH), 7.78(br s, 1H, NH), I = 6.0 Hz)
- NH), 7.78(br d, 1H, NH, *J* = 6.0 Hz).
  17: Colorless syrup. [α]<sub>0</sub><sup>27</sup> -21.6° (*c* 1.10, MeOH). <sup>1</sup>H NMR δ = 0.58(q, 6H, TES's CH<sub>2</sub>, *J* = 8.0 Hz), 0.89(t, 9H, TES's CH<sub>3</sub>, *J* = 8.0 Hz), 1.04(s, 9H, TPS's *t*-Bu), 3.60(s, 3H, oxazole's CH<sub>3</sub>), 3.76(s, 3H, OCH<sub>3</sub>), 4.01(m, 2H, Ser's β-H), 4.04(m, 1H, Ser's α-H), 4.79(m, 2H, Ser's β-H), 4.99(m, 1H, Ser's α-H), 5.15(ABq, 2H, Cbz's CH<sub>2</sub>, *J* = 12.3 Hz), 5.63(br s, 1H, NH), 7.31–7.62(m, 15H, Cbz's Ph and TPS's Ph × 2), 7.73(m, 1H, NH, *J* = 6.0 Hz).
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- 2: Colorless amorphous material.  $[α]_D^{26} 21.7^\circ$  (c 0.94, CHCl<sub>3</sub>). H NMR δ = 1.06, 1.09(each d, 6H, Val's CH<sub>3</sub> × 2, J = 6.5, 7.0 Hz), 1.52(d, 3H, Thr's CH<sub>3</sub>, J = 6.0 Hz), 1.60–1.75(m, 9H, Ip and propenyl's CH<sub>3</sub>), 2.30(m, 1H, Val's β-H), 2.61, 2.70(each s, 6H, oxazole's CH<sub>3</sub> × 2), 3.88(s, 3H, OCH<sub>3</sub>), 4.02(d, 1H, Thr's α-H, J = 7.5 Hz), 4.37(m, 1H, Thr's β-H), 4.54, 4.55(each d, 1H, Val's α-H, J = 6.5, 7.5 Hz), 5.15(m, 2H, Cbz's CH<sub>2</sub>), 5.45, 5.77, 5.95, 6.45, 6.56, 6.69(each s, 6H, vinyl's H), 6.56(br s, 1H, propenyl's H), 6.89(m, 1H, NH), 7.05–7.47(br s, 6H, Cbz's Ph and NH), 8.18(s, 1H, NH), 9.15(s, 1H, NH), 9.29(br s, 1H, NH).